Legiolert Test

Nonpotable water 1.0 mL protocol



The steps below are a guide only. Refer to the product insert (at idexx.com/waterinserts) for detailed instructions.

Chlorinated water must be collected in vessels with sodium thiosulfate.



 Add 100 mL of sterile diluent (e.g., deionized water, phosphate/ Butterfield's buffer, or 0.1% peptone) to a sterile vessel with no additives.



 Add contents of Legiolert* blister pack to the vessel containing 100 mL sterile diluent and shake until contents are dissolved. The solution may remain cloudy.



3. Add 2.0 mL of Legiolert pretreatment into a sterile tube.



4. Then add 2.0 mL of nonpotable water sample to the same tube.



 Start a 1 minute timer; then mix the contents. Keep the tube at room temperature for 60 seconds (±5 seconds). Mix the tube again before 60 seconds has elapsed.



6. At 1 minute, immediately transfer only 2.0 mL of the treated sample to the vessel containing the dissolved Legiolert reagent as prepared in steps 1 and 2.



 Cap the vessel and shake to dissolve. Sample may remain cloudy but can be added to the Quanti-Tray* when no large particles are visible.



 Agitate and then pour sample into a Quanti-Tray*/Legiolert* tray. Tap tray to remove any air bubbles from small wells.



9. Immediately seal the tray using the Quanti-Tray* Sealer PLUS.



10a. Place trays paper side down into humidified incubator at 37°C (±0.5°C) for 7 days. Provide adequate humidity to minimize Quanti-Tray water loss during 7 days. Do not remove trays from the incubator prior to the final read.



10b. Alternatively, insert tray(s) paper side down into an appropriately sized resealable plastic bag or box. Then incubate container at 37°C (±0.5°C) for 7 days. Do not remove trays from the incubator prior to the final read.



 Read results: Any brown and/or turbid wells are positive for Legionella pneumophila. Count positive wells and quantify using MPN table.

For technical support, contact IDEXX at 1-800-321-0207 or water@idexx.com.

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